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# Trichiol and 3-epitrichiol acetate, novel cytotoxic sterols with an unprecedented 2,6-dioxabicyclo[2.2.2]octan-3-one ring system from the myxomycete *Trichia favoginea* var. *persimilis*

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Abstract—Trichiol (1) and 3-epitrichiol acetate (2), two new sterols, have been isolated from field-collected fruit bodies of the myxomycete, *Trichia favoginea* var. *persimilis*, and their structures elucidated by spectral data. Trichiol (1) and 3-epitrichiol acetate (2) possess an unprecedented 2,6-dioxabicyclo[2.2.2]octan-3-one ring system. Trichiol (1) was cytotoxic against HeLa cells, while compound 2 proved to exhibit reversal effect against TNF-related apoptosis inducing ligand (TRAIL)-resistant Jurkat cell lines. © 2006 Elsevier Ltd. All rights reserved.

## 1. Introduction

In our search for bioactive natural products from myxomycetes,<sup>1,2</sup> we recently investigated a field-collected material of the fruit bodies of *Trichia favoginea* var. *persimilis* (Triciaceae) and reported the isolation of three new dibenzofurans, Kehokorins A–C.<sup>3</sup> Further investigation of the extract of this myxomycete has now resulted in isolation of two new sterols, trichiol (1), and 3-epitrichiol acetate (2), with a novel 2,6-dioxabicyclo[2.2.2]octan-3-one ring system. Here, we describe the isolation and structure elucidation of these sterols. Trichiol (1) was cytotoxic against HeLa cells, while 3-epitrichiol acetate (2) proved to exhibit reversal effect against TNF-related apoptosis inducing ligand (TRAIL)-resistant Jurkat cell lines.

## 2. Results and discussion

The fruit bodies of myxomycetes, *T. favoginea* var. *persimilis*,<sup>4</sup> collected in Kochi Prefecture, Japan, were

extracted with 90% MeOH and 90% acetone. The combined extracts were subjected to flash chromatography on silica gel and ODS, followed by further purification with reversed-phase HPLC (Develosil C30-UG-5) to give trichiol (1) and 3-epitrichiol acetate (2) in 0.01% and 0.02% yield, respectively.<sup>5</sup>

Trichiol (1)<sup>6</sup> showed a quasi-molecular ion peak at m/z 459 (M+H)<sup>+</sup> in its positive FAB mass spectrum, and its molecular formula was revealed as C<sub>29</sub>H<sub>46</sub>O<sub>4</sub> by the HRFABMS data [m/z 459.3429, (M+H)<sup>+</sup>,  $\Delta$  -4.6 mmu]. The IR absorption bands at 3419 and 1773 cm<sup>-1</sup> indicated the presence of hydroxy and carbonyl groups, and no particular UV absorption was observed for 1. The <sup>1</sup>H NMR spectrum of 1 in CDCl<sub>3</sub> (Table 1) showed signals for a dioxymethine proton at  $\delta_{\rm H}$  5.73 (1H, s), two oxymethine protons at  $\delta_{\rm H}$  3.59 (1H, tt, J = 9.5 and 4.8 Hz) and 4.09 (1H, dt, J = 6.8 and 1.5 Hz), and four methyl groups [one primary methyl at  $\delta_{\rm H}$  0.87 (3H, t, J = 7.0 Hz), two secondary methyls at  $\delta_{\rm H}$  0.82 (3H, d, J = 7.0 Hz), and at  $\delta_{\rm H}$  0.83 (3H, s)]. The <sup>13</sup>C NMR spectrum of 1 (Table 1) gave 29 signals assignable to one carbonyl ( $\delta_{\rm C}$  173.2), 2 sp<sup>3</sup> quaternary carbons ( $\delta_{\rm C}$  35.8 and 48.0), 11 sp<sup>3</sup> methines ( $\delta_{\rm C}$  100.4, 72.5, 71.2, 56.9, 55.2, 46.3, 45.0, 41.8, 36.6, 34.7, and 28.2), 11 sp<sup>3</sup> methylenes ( $\delta_{\rm C}$  38.0, 36.7, 35.7,

*Keywords*: Myxomycete; *Trichia favoginea* var. *persimilis*; Fruit body; 2,6-dioxabicyclo[2.2.2]octan-3-one ring; Cytotoxicity.

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Table 1. <sup>1</sup>H and <sup>13</sup>C NMR data of trichiol (1) and 3-epitrichiol acetate (2)

Positions	1 (CDCl <sub>3</sub> )		<b>2</b> (CDCl <sub>3</sub> )	
	$\delta_{\rm H} (J \text{ in Hz})$	$\delta_{ m C}$	$\delta_{\rm H} (J \text{ in Hz})$	$\delta_{ m C}$
1	(α) 0.99 m	36.7	1.23 m (2H)	32.7
	(β) 1.68 m			
2	(α) 1.80 m	31.4	(α) 1.72 m	26.0
	(β) 1.41 m		(β) 1.64 m	
3	3.59 tt (9.5, 4.8)	71.2	5.01 br s	70.0
4	(α) 1.56 m	38.0	1.46 m (2H)	32.8
	(β) 1.29 m			
5	1.13 m	45.0	1.47 m	40.2
6	1.22 m and 1.30 m	28.5	(α) 1.25 m	28.1
			(β) 1.18 m	
7	0.95 m and 1.73 m	32.4	1.00 m and 1.69 m	32.3
8	1.65 m	34.7	1.65 m	34.7
9	0.85 m	55.2	0.97 m	55.1
10		35.8		36.1
11	(α) 1.68 m	21.9	(α) 1.68 m	21.5
	(β) 1.38 m		(β) 1.36 m	
12	$(\alpha)$ 1.44 m	35.7	(α) 1.44 m	35.7
	(β) 2.36 m		(β) 2.37 m	
13		48.0		48.1
14	1.43 m	56.9	1.43 m	57.0
15	1.52 m (2H)	26.3	1.52 m (2H)	26.3
16	(α) 1.87 m	30.2	(α) 1.87 m	30.2
	(B) 1.60 m		(B) 1.60 m	
17	2.39 dd (10.8, 3.9)	36.6	2.40 dd (9.8, 3.9)	36.6
18	5.73 s	100.4	5.74 s	100.5
19	0.83 s (3H)	12.2	0.82 s (3H)	11.3
20	2.65 dd (3.9, 1.5)	46.3	2.66 dd (3.9, 1.5)	46.4
21		173.2		173.2
22	4.09 dt (6.8, 1.5)	72.5	4.09 dt (6.8, 1.5)	72.6
23	1.48 m and 1.68 m	31.7	1.49 m and 1.68 m	31.8
24	1.19 m	41.8	1.19 m	41.8
25	1.78 m	28.2	1.78 m	28.3
26	0.82 d (7.0) (3H)	18.7	0.82 d (7.0) (3H)	18.7
27	0.86 d (7.0) (3H)	18.9	0.86 d (7.0) (3H)	18.9
28	1.19 m and 1.36 m	22.7	1.19 m and 1.36 m	22.7
29	0.87 t (7.0) (3H)	11.9	0.87 t (7.0) (3H)	11.9
	× /× /		× /× /	170.6
			2.05 s (3H)	21.5

32.4, 31.7, 31.4, 30.2, 28.5, 26.3, 22.7, and 21.9), 4 sp<sup>3</sup> methyls ( $\delta_{\rm C}$  18.9, 18.7, 12.2, and 11.9). Since only one out of the seven unsaturation degrees was accounted for by the presence of a carbonyl group, 1 was inferred to contain six rings. The <sup>1</sup>H–<sup>1</sup>H COSY spectrum of 1 revealed the following five partial structures: (i) from H-1 to H-7, (ii) from H-8 to H-12 and H-14, (iii) from H-15 to H-20, (iv) from H-22 to  $H_3$ -29, and (v) from H-25 to H<sub>3</sub>-27 (Fig. 1). Evidence for the connections of these partial structures was provided by the HMBC <sup>1</sup>H-<sup>13</sup>C long-range connectivity data (Fig. 1). The presence of a 2,6-dioxabicyclo[2.2.2]octan-3-one ring was suggested by the following HMBC correlations: H-17/C-21, H-18/C-13, H-18/C-17, H-18/C-21, H-18/C-22, H-20/C-13, H-20/C-17, H-20/C-21, H-22/C-17, and H-22/C-21. HMBC correlations for H-23/C-20, H-23/C-22, H-23/ C-24, H-23/C-25, H-23/C-28, H<sub>3</sub>-26/C-24, H<sub>3</sub>-26/C-25, H<sub>3</sub>-26/C-27, H<sub>3</sub>-27/C-25, H<sub>3</sub>-27/C-26, H<sub>2</sub>-28/C-23, H<sub>2</sub>-28/C-24, H<sub>2</sub>-28/C-25, H<sub>2</sub>-28/C-29, H<sub>3</sub>-29/C-24, and H<sub>3</sub>-29/C-28 indicated the existence of a 2-ethyl-3-methylbutyl side chain (C-23 to C-29) attached on C-22. Thus, the planar structure of trichiol was revealed as 1. The relative stereochemistry of trichiol (1) was eluci-



Figure 1. Key  ${}^{1}H{-}^{1}H$  COSY and HMBC Data of 1.

dated by a combination of observed coupling constants and NOESY data (Fig. 2). The NOESY correlations observed for H<sub>3</sub>-19/H-2 $\beta$ , H<sub>3</sub>-19/H-4 $\beta$ , H-3/H-2 $\alpha$ , H-3/H-4 $\alpha$ , and H-3/H-5, as well as the coupling constants observed for H-3 ( $J_{3,2\beta} = J_{3,4\beta} = 9.8$  Hz and  $J_{3,2\alpha} = J_{3,4\alpha} = 4.8$  Hz) suggested that the A-ring exists in a chair conformation, and the secondary hydroxy



Figure 2. Key NOESY Data of 1.

group on C-3 was oriented in a β-equatorial arrangement, while the tertiary methyl group on C-10 (C-19) was in a  $\beta$ -axial position. The NOESY cross-peaks observed for H-8/H<sub>3</sub>-19, H-8/H-18, H-9/H-1α, H-9/ H-5, H-12α/H-17, H-17/H-14, H-18/H-11β implied that the B- and C-rings exist in a chair conformation, and the C/D rings were fused with a *trans*-orientation and the H-18 (C-18 methine carbon) was therefore located in the  $\beta$ -position. Since NOESY correlations were observed for H-20/H-16β, H-17/H-12α, H-17/H-14, and H-17/H-16a, the C-20 methine carbon was revealed to have  $\beta$ -position. The presence of cross-peaks between H-12 $\beta$ /H-23 ( $\delta_{\rm H}$  1.68) and H-17/H-23 ( $\delta_{\rm H}$  1.48) provided evidence that the C-23 side chain was in the  $\alpha$ -position and the H-22 in the  $\beta$ -position. The configuration of the C-24 chiral center bearing the ethyl group remained undetermined due to conformational lability. From several species of myxomycetes such as *Physarum*<sup>7</sup> and Didymium,<sup>8</sup> clionasterol (3) was isolated as a major sterol, and 3 was a C-24 epimer of  $\beta$ -sitosterol (4) which is known as a major plant sterol.9

The molecular formula of 3-epitrichiol acetate (2)<sup>10</sup> was revealed as  $C_{31}H_{48}O_5$  by the HRFABMS data [m/z501.3557,  $(M+H)^+$ ,  $\Delta$  –2.3 mmu], having a  $C_2H_2O$  unit more than that of trichiol (1). The <sup>1</sup>H and <sup>13</sup>C NMR spectral data of 2 (Table 1) were parallel to those of compound 1, except for the fact that the NMR signal due to a methyl group was observed at  $\delta_H$  2.05 (3H, s) and an oxymethine proton on C-3 resonated at relatively lower field for 2 (H-3,  $\delta_H$  5.01). Although analysis of the <sup>1</sup>H–<sup>1</sup>H COSY, HMQC, HMBC, and NOESY data of 2 showed that the planar structure of 2 corresponded to



an acetate of compound 1, the acetoxy-bearing methine proton (H-3) appeared as a broad singlet; this observation suggested that this proton (H-3) has small coupling constants with vicinal methylene protons on C-2 and C-4, implying  $\beta$ -equatorial orientation of H-3. Thus, compound 2 was inferred to be 3-epitrichiol acetate [= an acetate of C-3 epimer of trichiol (1)], which was further supported by the following findings. Trichiol (1) was treated with acetic anhydride and pyridine to give its acetate (5), FABMS m/z 501 (M+H)<sup>+</sup>.<sup>11</sup> The <sup>1</sup>H NMR data of 5 were almost parallel but not identical with those of compound 2. The H-3 of 5, having  $\alpha$ -axial configuration, resonated at relatively higher field ( $\delta_{\rm H}$ 4.69) as a triplet of triplet ( $J_{3,2\beta} = J_{3,4\beta} = 10.6$  Hz and  $J_{3,2\alpha} = J_{3,4\alpha} = 5.4$  Hz), while H-3 of 2 was observed as a broad singlet at  $\delta_{\rm H}$  5.01 (vide supra).

Trichiol (1) and 3-epitrichiol acetate (2) belong to a new class of sterols possessing a 2,6-dioxabicyclo[2.2.2]-octan-3-one ring system. This unique ring system may be generated, from a sterol such as 3 or 4, through oxidation of the C-18 methyl into an aldehyde, the C-21 methyl into a carboxyl group, and the C-22 methylene into a hydroxyl-bearing methine carbon, followed by formation of a lactone and acetal functionalities (Scheme 1). Although several types of sterols with



Scheme 1. A conceivable biogenetic pathway of 1.

chemically-unique structures had been isolated from a variety of natural sources,<sup>12</sup> the 2,6-dioxabicyclo[2.2.2]octan-3-one ring system is so far unknown as a partial structure, to the best of our knowledge. Trichiol (1) exhibited cell growth inhibitory activity against the HeLa human epithelial carcinoma cell line (>90% inhibition at 25  $\mu$ M, IC<sub>50</sub> 12.5–25  $\mu$ M), while compound **2** and clionasterol (**3**)<sup>13</sup> were almost inactive (IC<sub>50</sub> > 25 $\mu$ M). On the other hand, compound **2** proved to exhibit reversal effect against TNF-related apoptosis inducing ligand (TRAIL)-resistant Jurkat cell lines.<sup>14</sup> Although 50  $\mu$ M of compound **2** and 0.125  $\mu$ g/mL of TRAIL alone did not show growth inhibition on Jurkat cells, the combination of these compounds produced a remarkable inhibitory activity on the growth of Jurkat cells (T/C% = 15.8).

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- 4. The fruit bodies of *Trichia favoginea* var. *persimilis* were collected at Tosayamada-shi in Kochi Prefecture, Japan, in January 2004. A voucher specimen (#25634) is maintained by Y.Y. (Ohtsu-ko, Kochi).
- 5. The wild fruit bodies (17.6 g) were extracted with 90% MeOH (200 mL  $\times$  3) and 90% acetone (200 mL  $\times$  1). The combined MeOH and acetone extracts (1.0 g) were subjected to silica gel column chromatography (30  $\times$  180 mm) eluted with 0–100% methanol in chloroform. The fraction eluted with 20% MeOH in CHCl<sub>3</sub> afforded arcyriaflavin C (40 mg),<sup>15,16</sup> and the fraction (86 mg) eluting with 100% CHCl<sub>3</sub> was separated by the second silica gel chromatography (15  $\times$  280 mm) eluted with 0–10% acetone in chloroform, and the fraction (10 mg) eluting with 5–10% acetone in CHCl<sub>3</sub> followed by ODS column (MeOH/H<sub>2</sub>O, 3:1) to afford trichiol (1, 2.4 mg). The fraction of the first silica gel column (52 mg) eluting with 100% CHCl<sub>3</sub> was

separated by the second ODS column (MeOH/H<sub>2</sub>O, 95:5), followed by further purification with reversed-phase HPLC (Develosil C30-UG-5, 90–100% MeOH; ODS-UG-5, 60% CH<sub>3</sub>CN) to afford 3-epitrichiol acetate ( $\mathbf{2}$ , 3.1 mg).

- 6. Trichiol (1): amorphous solid;  $[\alpha]_D^{22} +98$  (c 0.5, CHCl<sub>3</sub>); IR (film)  $\nu_{max}$  3419 and 1773 cm<sup>-1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR (Table 1); FABMS m/z 459 (M+H)<sup>+</sup>; HRFABMS calcd for C<sub>29</sub>H<sub>46</sub>O<sub>4</sub> (M+H)<sup>+</sup> 459.3474, found m/z 459.3429.
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- 10. 3-Epitrichiol acetate (**2**): amorphous solid;  $[\alpha]_{D}^{22}$  +90 (*c* 0.5, CHCl<sub>3</sub>); IR (film)  $\nu_{max}$  1775 and 1733 cm<sup>-1</sup>; <sup>1</sup>H and <sup>13</sup>C NMR (Table 1); FABMS *m/z* 501 (M+H)<sup>+</sup>; HRFABMS calcd for C<sub>31</sub>H<sub>49</sub>O<sub>5</sub> (M+H)<sup>+</sup> 501.3580, found *m/z* 501.3557.
- 11. Trichiol (1, 0.4 mg) was treated with Ac<sub>2</sub>O/pyridine (1:1, 0.2 mL) overnight at room temperature. The solvent was then evaporated and the residue purified by silica gel chromatography (hexane/EtOAc, 8:2), followed by Sephadex LH-20 column (CHCl<sub>3</sub>/MeOH, 1:1) to afford compound **5** (0.5 mg): <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta_{\rm H}$  4.69 (1H, tt, J = 10.6 and 5.4 Hz; H-3), 5.72 (1H, s; H-18), 4.09 (1H, dt, J = 7.2 and 1.5 Hz; H-22), 2.65 (1H, dd, J = 4.2 and 1.5 Hz; H-20), 2.40 (1H, dd, J = 9.6 and 4.2 Hz; H-17), and 2.02 (3H, s; CH<sub>3</sub>CO); FABMS m/z 501 (M+H)<sup>+</sup>.
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- 13. Clionasterol (3) was isolated from cultured *Didymium minus* previously in our laboratory.<sup>8</sup>
- 14. For Jurkat cells,  $3.5 \times 10^5$  cells/mL of the cells were seeded in 95 µL of culture medium per well in 96-well microtiter plates, and were treated with 5 µL of graded concentrations of samples in the absence or presence of 0.125 µg/mL of TRAIL, and were then incubated for 42 h at 37 °C in a 5% CO<sub>2</sub>-95% air atmosphere. Cell viability was determined by the colorimetric assay using alamer blue: Fields, R. D.; Lancaster, M. V. *Am. Biotechnol. Lab.* **1993**, *11*, 48-50.
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